## APPENDIX N BULK MILK TANKER SCREENING TEST FORM

# CHARM® FLUSLBL (Raw Commingled Cow Milk) FLUNIXIN and BETA-LACTAM TEST IMS #9-C16

[Unless otherwise stated all tolerances are ±5%]

## GENERAL REQUIREMENTS

1. See Appendix N General Requirements (App. N GR) items 1-8 & 15

## SAMPLES

#### 2. See App. N GR item 9

#### **APPARATUS & REAGENTS**

#### 3. Equipment

- a. Charm Sciences Strip Incubator: 56±1°C 8 min timer 56±1°C Charm EZ display when message 'Add milk to strip and close door'
  - 1. Clean, properly maintained and located on a level surface
  - 2. Check temperature daily (day of use); maintain records
    - a. Charm EZ printout acceptable for daily temperature check (annual accuracy check required); maintain records
  - 3. Temperature measuring device for each incubator (App N. GR item 3)
  - 4. Lid closed (slightly sprung so that timer not active) when not running tests
  - 5. Incubator Temperature: \_\_\_\_\_
- b. ROSA® Reader, ROSA Pearl Reader (with or without ROSA Barcode option), Charm EZ or Charm Sciences equivalent with print out or download of data; manual available

| Serial Number: |  |
|----------------|--|
|----------------|--|

|     | <ol> <li>ROSA Reader V1.07 or higher<br/>(or if ROSA Pearl Reader or Charm EZ see 3.b.2)</li> </ol>   |   |  |  |  |  |  |
|-----|---|---|--|--|--|--|--|
|     |   | a. Calibrators  |  |  |  |  |  |
|     |   | Range(s) Result   |  |  |  |  |  |
|     |   | Low:  |  |  |  |  |  |
|     |   | High:   |  |  |  |  |  |
|     |   | b. Maintain records   |  |  |  |  |  |
|     | 2.  | ROSA Pearl Reader V3.00 or higher or Charm EZ               |  |  |  |  |  |
|     |   | a. Calibrators - Low and High for use in all assay channels |  |  |  |  |  |
|     |   | Range(s) Result   |  |  |  |  |  |
|     |   |   |  |  |  |  |  |
|     |   | Low: (darker magenta)                                       |  |  |  |  |  |
|     |   | High: (lighter pink)  |  |  |  |  |  |
|     |   |   |  |  |  |  |  |
|     |   | b. Maintain records   |  |  |  |  |  |
|     | 3.  | Calibrator serial numbers match ROSA reader SN              |  |  |  |  |  |
|     | 4. <b>Do not proceed if out of range.</b> Manufacturer should be contacted for corrective actions   |   |  |  |  |  |  |
|     | 5. Printer or computer link for hardcopy download   |   |  |  |  |  |  |
| C.  | Pipettor - 300 μL and disposable tips (see App. N GR item 7)  |   |  |  |  |  |  |
| d.  | Or single use 300 µL ROSA-pipet with overflow bulb to accurately measure amount of sample; supplied by manufacturer <b>(screening only)</b> |   |  |  |  |  |  |
| Rea | agen  | uts   |  |  |  |  |  |
| a.  | Tes   | st Strips (EZ Compatible for Charm EZ)                      |  |  |  |  |  |
|     | Lot   | t #: Exp. Date:   |  |  |  |  |  |
|     | QC Date: By:  |   |  |  |  |  |  |

4.

| b.  | Positive Control (labeled as 2 ppb Flunixin and 5 ppb Penicillin G<br>Standard)                                |  |  |  |  |  |  |  |
|-----|--|--|--|--|--|--|--|--|
|     | 1.   | 5 ppb Penicillin G Standard                    |  |  |  |  |  |  |
|     |  | Lot #: Exp. Date:                              |  |  |  |  |  |  |
|     | 2.   |  | bb Lyophilized Flunixin  |  |  |  |  |  |
|     |  | Lot #: Exp. Date:                              |  |  |  |  |  |  |
|     | 3.   |  |  |  |  |  |  |  |
|     | 4.   | Prep   | paration   |  |  |  |  |  |
|     | a. Add 10.0 mL negative raw milk (item 5.d) to Flunixin control (item 4.b.2), and allow to rehydrate for 5 min |  |  |  |  |  |  |  |
|     |  | b.   | Add 8.0 mL of reconstituted Flunixin control (item 4.b.4.a) to<br>Positive Control (item 4.b.1), and allow to rehydrate<br>for 5 min                           |  |  |  |  |  |
|     |  | C.   | Or, alternative to 4.b.4.a and 4.b.4.b, add 5.0 mL negative milk (item 5.d) to Flunixin and Penicillin G tablet (item 4.b.3), and allow to rehydrate for 5 min |  |  |  |  |  |
| C.  | Neg  | jative Control                                 |  |  |  |  |  |  |
|     | 1.   | Previously negative tested raw milk (item 5.d) |  |  |  |  |  |  |
| Rea | Reagent stability  |  |  |  |  |  |  |  |
| a.  | FLUSLBL reagents received refrigerated   |  |  |  |  |  |  |  |
| b.  | Store reagents at 0.0-4.5°C, desiccant blue, maintain no longer than manufacturer's expiration date            |  |  |  |  |  |  |  |
|     | 1.   | Do r   | not use if desiccant indicator is white or pink  |  |  |  |  |  |
| C.  |  |  | Control - Manufacturer supplied; maintain no longer than<br>urer's expiration date   |  |  |  |  |  |
|     | 1.   |  | onstituted Control (4.b.4), tested +400 or more positive; use<br>in 48 hours when maintained at 0.0-4.5°C  |  |  |  |  |  |
|     | Lab Prep. Date: Lab Exp. Date:   |  |  |  |  |  |  |  |

5.

|    | <ol> <li>Or, aliquot within 24 hours and freeze at =15°C or colder in a<br/>non-frost-free freezer or in an insulated foam container in a<br/>frost-free freezer; use within 3 weeks</li> </ol> |   |  |  |  |  |
|----|---|---|--|--|--|--|
|    |   | Lab Date prep: Lab Exp. Date:   |  |  |  |  |
|    |   | a. Thaw slowly overnight in refrigerator or more rapidly in cold water. Mix well until sample is homogeneous  |  |  |  |  |
|    |   | 1. Do not use if there is visible protein precipitation   |  |  |  |  |
|    |   | b. Store at 0.0-4.5°C and use within 24 hours; do not refreeze  |  |  |  |  |
|    | 3.  | Day of use, must produce +400 or greater reading; maintain records  |  |  |  |  |
|    |   | Test Value:   |  |  |  |  |
|    |   | Do not proceed if out of range  |  |  |  |  |
| d. | Neg   | ative Control - raw milk tested –400 or more negative with FLUSLBL test   |  |  |  |  |
|    | Sam   | nple ID: Test Value:  |  |  |  |  |
|    | Date  | e tested:   |  |  |  |  |
|    | 1. Used within 72 hours when maintained at 0.0-4.5°C  |   |  |  |  |  |
|    | 2.  | Or, aliquot within 24 hours and freeze at –15°C or colder in a non-frost-free freezer or in an insulated foam container in a frost-free freezer; use within 3 weeks |  |  |  |  |
|    |   | Lab Prep. Date: Lab Exp. Date:  |  |  |  |  |
|    |   | a. Thaw slowly overnight in refrigerator or more rapidly in cold water.<br>Mix well until sample is homogeneous   |  |  |  |  |
|    |   | 1. Do not use if there is visible protein precipitation   |  |  |  |  |
|    |   | b. Store at 0.0-4.5°C and use within 24 hours; do not refreeze  |  |  |  |  |
|    | 3.  | Day of use must produce –400 or more negative with FLUSLBL test:  |  |  |  |  |
|    |   | Do not proceed if out of range  |  |  |  |  |

# TECHNIQUE

| 6. | Dai                               | Daily Performance and Operation Check  |  |  |  |  |  |  |
|----|-----------------------------------|--|--|--|--|--|--|--|
|    | a.                                | . See App. N GR items 10.b-d   |  |  |  |  |  |  |
|    | b.                                | If using ROSA reader Version 1.07 and higher, or ROSA Pearl, use<br>ESC 5 reader function to enter performance monitoring mode of reader; if<br>using Charm EZ, use Menu to enter Performance Monitoring mode and<br>"Perf Mon" to enter daily performance check; refer to manual for directions |  |  |  |  |  |  |
|    | C.                                | Check Calibrators, items 3.b.1 & 3.b.2   |  |  |  |  |  |  |
|    | d.                                | Positive and negative controls must give appropriate readings prior to any sample analysis (see App. N GR item 10.a)   |  |  |  |  |  |  |
|    | e.                                | Controls in-range when in performance monitoring mode, ROSA reader version 1.07 and higher, ROSA Pearl or Charm EZ   |  |  |  |  |  |  |
|    |                                   | 1. If out of range, manufacturer should be contacted for corrective action, 800-343-2170   |  |  |  |  |  |  |
|    | f. Do not proceed if out of range |  |  |  |  |  |  |  |
| 7. | Tes                               | t Procedure  |  |  |  |  |  |  |
|    | a.                                | Set out required number of test strips for samples to be tested in one day and place them in a dry labeled container at room temperature, or take out strips as needed   |  |  |  |  |  |  |
|    |                                   | 1. Discard unused test strips at the end of the day  |  |  |  |  |  |  |
|    | b.                                | Label test strips, one for each test sample and each control. Avoid crushing sample compartment(s)   |  |  |  |  |  |  |
|    | C.                                | vortex for 10 sec at maximum setting; use within 3 min (samples/controls must be in appropriate containers to allow the use of vortexing)  |  |  |  |  |  |  |
|    | d.                                |  |  |  |  |  |  |  |
|    | e.                                | While holding strip flat, peel back plastic (to 'peel to here' line) to expose sample pad compartment. Avoid lifting the wick and sponge under tape  |  |  |  |  |  |  |
|    |                                   | <ol> <li>For multiple samples, complete steps 7.d-g for each sample/control,<br/>before starting test of next sample</li> </ol>  |  |  |  |  |  |  |
|    |                                   | 2. Complete all samples within 2 min of placing first strip in incubator   |  |  |  |  |  |  |

| f. | Add 300 $\mu$ L of mixed sample/control to corresponding strip |  |  |  |  |  |  |
|----|--|--|--|--|--|--|--|
|    | 1.   |  |  |  |  |  |  |
|    |  | a. Remove tip from liquid  |  |  |  |  |  |
|    |  | b.   |  | le holding the pipettor vertically, expel test portion slowly either side well of appropriate strip  |  |  |  |
|    | 2.   | Using new manufacturer-provided ROSA-pipet (item 3.d) for each control/sample <b>[Screening only]</b>  |  |  |  |  |  |
|    |  | a.   | rese<br>and                                      | ress top bulb while holding vertically with bulb and overflow<br>rvoir side pointing down, draw up test portion avoiding foam<br>bubbles. Sample should completely fill pipet shaft and<br>flow into the bottom half of the overflow reservoir |  |  |  |
|    |  | b.   | Rem  | nove tip from liquid   |  |  |  |
|    |  | C.   | slow   | le holding the ROSA-pipet vertically, expel test portion<br>ly into either side well of appropriate strip. Excess portion<br>uld remain in reservoir   |  |  |  |
| g. | Re-s   | seal p   | eal plastic firmly around sample pad compartment |  |  |  |  |
| h. | ROS  | ROSA Reader and Charm EZ (read only mode)  |  |  |  |  |  |
|    | 1.   | Close lid and latch ROSA incubator to start automatic timer in the incubator. If no automatic timer in incubator, set external timer for 8 min not to exceed 9 min                     |  |  |  |  |  |
|    | 2.   | At end of incubation visually inspect C (Control) line. An absent C line, a partial C line or an indistinct C line indicates an invalid test, and the sample/control must be re-tested |  |  |  |  |  |
|    | 3.   | Insert only valid test(s) in reader  |  |  |  |  |  |
|    |  | a.   | ROS  | SA reader set to appropriate channel   |  |  |  |
|    |  |  | 1.   | Press ENTER; reading and interpretation appear in 5 sec,<br>read strips within 5 min of completion of incubation. Strips<br>may be held vertically, sample compartment down while<br>waiting to be read  |  |  |  |

|     |  |    | b.   | Cha<br>inse | rm EZ automatically sets channel when color coded strip<br>rted   |  |  |
|-----|--|----|------|-------------|---|--|--|
|     |  |    |      | 1.          | Close door; reading and interpretation appear in 5 sec,<br>read strips within 5 min of completion of incubation. Strips<br>may be held vertically, sample compartment down while<br>waiting to read |  |  |
|     | i. Charm EZ (incubate and read mode)   |    |      |             |   |  |  |
|     | 1. Charm EZ automatically sets channel and incubator temperature when color coded strip inserted. Optionally, enter sample ID  |    |      |             |   |  |  |
|     |  | 2. | Peel | strip       | (7.e) and add milk (7.f)  |  |  |
|     |  | 3. | Clos | e do        | or to begin   |  |  |
|     |  | 4. | Cha  | rm Ež       | Z automatically prompts for further testing when positive _   |  |  |
| 8.  | Interpretation with ROSA Reader  |    |      |             |   |  |  |
|     | <ul> <li>a. If there is a negative or zero reading on the reader, sample is a<br/>Negative (NF)</li> </ul>   |    |      |             |   |  |  |
|     | <ul> <li>b. If there is a positive reading on the reader, sample is an<br/>Initial Positive</li> </ul>   |    |      |             |   |  |  |
| 9.  | Verification of Initial Positive Tanker Samples (see App. N GR item 11);<br>Confirmation of Presumptive Positive Tanker Samples (see App. N GR<br>item 12); and Traceback of Producer(s) on a Confirmed Positive Tanker<br>(see App. N GR item 13) |    |      |             |   |  |  |
| 10. | . Reporting (see App. N GR item 14)  |    |      |             |   |  |  |

8.

9.